

PATENT COOPERATION TREATY

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

From the INTERNATIONAL BUREAU

To:

Commissioner
US Department of Commerce
United States Patent and Trademark
Office, PCT
2011 South Clark Place Room
CP2/5C24
Arlington, VA 22202
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year)

14 May 2001 (14.05.01)

International application No.

PCT/IB00/01276

Applicant's or agent's file reference

7589/2053 TS

International filing date (day/month/year)

08 September 2000 (08.09.00)

Priority date (day/month/year)

09 September 1999 (09.09.99)

Applicant

GHISALBERTI, Carlo

1. The designated Office is hereby notified of its election made:



in the demand filed with the International Preliminary Examining Authority on:

12 March 2001 (12.03.01)



in a notice effecting later election filed with the International Bureau on:

2. The election ☒ was

was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland

Facsimile No.: (41-22) 740.14.35

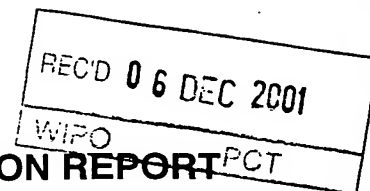
Authorized officer


Olivia TEFY

Telephone No.: (41-22) 338.83.38

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)



Applicant's or agent's file reference 7589/2053 TS		FOR FURTHER ACTION See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/IB00/01276	International filing date (day/month/year) 08/09/2000	Priority date (day/month/year) 09/09/1999	
International Patent Classification (IPC) or national classification and IPC C09B61/00			
Applicant GHISALBERTI, Carlo			
<p>1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.</p> <p>2. This REPORT consists of a total of 7 sheets, including this cover sheet.</p> <p><input type="checkbox"/> This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).</p> <p>These annexes consist of a total of sheets.</p>			
<p>3. This report contains indications relating to the following items:</p> <ul style="list-style-type: none"> I <input checked="" type="checkbox"/> Basis of the report II <input type="checkbox"/> Priority III <input type="checkbox"/> Non-establishment of opinion with regard to novelty, inventive step and industrial applicability IV <input type="checkbox"/> Lack of unity of invention V <input checked="" type="checkbox"/> Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement VI <input type="checkbox"/> Certain documents cited VII <input checked="" type="checkbox"/> Certain defects in the international application VIII <input checked="" type="checkbox"/> Certain observations on the international application 			
Date of submission of the demand 12/03/2001		Date of completion of this report 04.12.2001	
Name and mailing address of the international preliminary examining authority:  European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465		Authorized officer Stroeter, T Telephone No. +49 89 2399 8088	



INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/IB00/01276

I. Basis of the report

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17):*)

Description, pages:

1-32 as originally filed

Claims, No.:

1-16 as originally filed

Drawings, sheets:

1/6-6/6 as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
- ☐ the claims, Nos.:

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/IB00/01276

☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Yes:	Claims 3-5,7-13
	No:	Claims 1,2,6,14-16
Inventive step (IS)	Yes:	Claims
	No:	Claims 1-16
Industrial applicability (IA)	Yes:	Claims 1-16
	No:	Claims

2. Citations and explanations
see separate sheet

VII. Certain defects in the international application

The following defects in the form or contents of the international application have been noted:
see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:
see separate sheet

Re Item V

Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

The following statements concerning examination according to Article 33 PCT are to be seen as preliminary with respect to the objections that have been raised under VIII due to lacking clarity of the present application.

1 Prior art documents

Reference is made to the following documents. The given numbering will be adhered to in the rest of the procedure:

- D1: US-A-5 384 116 (ORLOW SETH J ET AL)
- D2: FR-A-2 704 554 (OREAL)
- D3: DE 197 17 837 A (KNOERLE RAINER DR ET AL.)
- D4: WO 96 25920 A (UNIV YALE) cited in the application
- D5: PATENT ABSTRACTS OF JAPAN vol. 015, no. 243 (C-0842), 21 June 1991
& JP 03 077813 A (RINJIRO SARUNO)
- D6: US-A-5 122 461 (LEE FENG-TSUN ET AL)
- D7: WO 92 00373 A (BIOSOURCE GENETICS CORP)
- D8: EP-A-0 363 792 (BIOSOURCE GENETICS CORP)
- D9: PATENT ABSTRACTS OF JAPAN vol. 006, no. 078 (C-102), 15 May 1982
& JP 57 014517 A (SANSHO SEIYAKU KK)
- D10: WO 93 23480 A (INTERCAST EUROP SPA ;GALLAS JAMES (US))
- D11: WO 94 12644 A (BIOSOURCE GENETICS CORP)

D1, D2, D4, D5, D7, D8 and D10 are directed to the provision of synthetic melanins. D3 is directed to the investigation of the melanin affinity of substances via HPLC. D6 reveals the preparation of pyrocatecholic compounds while avoiding the formation of melanins. D9 is directed to quercetine as an additive in (i. a. melanin-containing) cosmetics. Finally, D11 discloses a method for making extracellular tyrosinase.

2 Novelty (Article 33(2) PCT) and Inventive step (Article 33(3) PCT)

The melanins claimed in present **claim 1** are not novel due to the following reasoning: Melanins prepared starting from "plant polyphenols" as monomers are already described in the prior art as can be taken from D1 (examples 2,3), D2 (example 1), D3 (page 2, line 43-53), D4 (example 1) and D10 (claim 4). It is to be noted, that the "Red:Green:Blue ratio" can at present not be seen as a distinguishing feature since the relevant prior art documents mentioned do not contain such technical feature to allow a comparison. It is the Applicant's burden to prove that the present melanins have different ratios compared with those of the prior art.

Present **claim 2** is not novel, since D4 cites e.g. 3,4-dihydroxybenzoic acid (precursor E) which falls under the scope of present formula (I). Present **claim 6** is not novel, too, since D3 discloses L-DOPA as a monomer for the melanin described therein.

The subject-matter of product **claims 3-5** is novel due to the selection of certain plant polyphenols. However, this selection does not define an inventive contribution in view of closest prior art D1.

Method **claims 7-13** appear to be novel but only differ from the procedures laid down in D1 via minor modifications and as such do not involve an inventive step either.

The compositions of **claims 14-16** are not novel as far as they contain compounds of claims 1 or 2 which are known and which already have been used in such compositions, see D1, claim 1 and D4, examples.

Summary: The present application does not fulfil the requirements of Articles 33(2) and (3) PCT since the subject-matter of present claims 1, 2, 6 and 14-16 is not novel and that of claims 3-5 and 7-13 not inventive.

Thus, if the Applicant is able to restore novelty to the present claims he is also requested to prove an advantageous effect of the present melanins vis-à-vis those of the most relevant prior art D1.

3 Industrial applicability (Article 33(4) PCT)

The subject-matter of the present set of claims 1 to 16 is in accordance with the requirements of Article 33(4) PCT.

Re Item VII

Certain defects in the international application

To meet the requirements of Rule 5.1(a)(ii) PCT, the document D1 should be identified in the description and the relevant background art disclosed therein should be briefly discussed.

Re Item VIII

Certain observations on the international application

1 Clarity of the claims (Article 6 PCT)

(1) Present claim 1 should be rendered clear by specifying the expressions "plant polyphenol" and "eumelanin precursor" according to the content of the description. Furthermore, in this context, it is noted that "eumelanin precursors" as L-DOPA do fall under the definition given for "plant polyphenols" and as such melanins are comprised in the scope of said claim which are prepared from only one starting compound.

(2) Formula (IV) of claim 2 is missing a substituent attached to the C=R³ group.

(3) Claims 3-6 refer to claims 1 and 2 but should only refer to claim 2 since formula (I) is first introduced in claim 2.

(4) Claim 9 is formulated to be dependent on claim 7 but specifies a "chemical oxidizing agent" which is not mentioned in claim 7.

2 Clarity of the description (Article 5 PCT)

(1) Line 18 on page 2 should read "...L-tyrosine to L-dopa..."

(2) To avoid confusion, tables II, III, IV, V and again III on pages 26, 27, 28 and 31 should be renamed in view of the tables I-IV on pages 21, 22, 24 and 25.

(3) Figure 2 of the present application should be verified by the Applicant since the lower four molecules comprise a =NH- group (charge?).

PATENT COOPERATION TREATY

PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference 7589/2053 TS	FOR FURTHER ACTION see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
International application No. PCT/IB 00/ 01276	International filing date (day/month/year) 08/09/2000	(Earliest) Priority Date (day/month/year) 09/09/1999
Applicant GHISALBERTI, Carlo		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.



It is also accompanied by a copy of each prior art document cited in this report.

1. Basis of the report

- a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.



the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

- b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :



contained in the international application in written form.



filed together with the international application in computer readable form.



furnished subsequently to this Authority in written form.



furnished subsequently to this Authority in computer readable form.



the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.



the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. ☐ **Certain claims were found unsearchable** (See Box I).

3. ☐ **Unity of invention is lacking** (see Box II).

4. With regard to the **title**,



the text is approved as submitted by the applicant.



the text has been established by this Authority to read as follows:

5. With regard to the **abstract**,



the text is approved as submitted by the applicant.



the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the **drawings** to be published with the abstract is Figure No.



as suggested by the applicant.



because the applicant failed to suggest a figure.



because this figure better characterizes the invention.



None of the figures.

INTERNATIONAL SEARCH REPORT

International Application No

PC 00/01276

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C09B61/00 C09B69/10 A61K7/42

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C09B A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 384 116 A (ORLOW SETH J ET AL) 24 January 1995 (1995-01-24) examples	1,2,6, 14,16
X	FR 2 704 554 A (OREAL) 4 November 1994 (1994-11-04) page 7, line 23 - line 25	1,7
X	DE 197 17 837 A (KNOERLE RAINER DR ;FEUERSTEIN THOMAS DR (DE); LIMBERGER NORBERT DR) 29 October 1998 (1998-10-29) claims	1
X	WO 96 25920 A (UNIV YALE) 29 August 1996 (1996-08-29) cited in the application examples	1,2,6, 14,16

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

G document member of the same patent family

Date of the actual completion of the international search

21 December 2000

Date of mailing of the international search report

02/01/2001

Name and mailing address of the ISA

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Authorized officer

Ketterer, M

INTERNATIONAL SEARCH REPORT

International Application No

PC 8 00/01276

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	PATENT ABSTRACTS OF JAPAN vol. 015, no. 243 (C-0842), 21 June 1991 (1991-06-21) & JP 03 077813 A (RINJIRO SARUNO), 3 April 1991 (1991-04-03) abstract ----	1, 14, 15
A	US 5 122 461 A (LEE FENG-TSUN ET AL) 16 June 1992 (1992-06-16) column 1, line 1 -column 2, line 18; claims ----	1, 7
A	WO 92 00373 A (BIOSOURCE GENETICS CORP) 9 January 1992 (1992-01-09) examples ----	1, 7
A	EP 0 363 792 A (BIOSOURCE GENETICS CORP) 18 April 1990 (1990-04-18) examples ----	1, 7
A	PATENT ABSTRACTS OF JAPAN vol. 006, no. 078 (C-102), 15 May 1982 (1982-05-15) & JP 57 014517 A (SANSHO SEIYAKU KK), 25 January 1982 (1982-01-25) abstract ----	1
A	WO 93 23480 A (INTERCAST EUROP SPA ;GALLAS JAMES (US)) 25 November 1993 (1993-11-25) claims ----	1
A	WO 94 12644 A (BIOSOURCE GENETICS CORP) 9 June 1994 (1994-06-09) examples -----	1, 7, 14-16

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PC 00/01276

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 5384116 A	24-01-1995	US 5227459 A US 5225435 A US 5218079 A AU 3977393 A EP 0638101 A JP 7509012 T WO 9321252 A CA 2092205 A EP 0548110 A JP 6505960 T WO 9216189 A US 5618519 A	13-07-1993 06-07-1993 08-06-1993 18-11-1993 15-02-1995 05-10-1995 28-10-1993 26-04-1992 30-06-1993 07-07-1994 01-10-1992 08-04-1997
FR 2704554 A	04-11-1994	CA 2139115 A DE 69422123 D DE 69422123 T EP 0647255 A WO 9425531 A JP 7508554 T US 5776241 A	10-11-1994 20-01-2000 06-04-2000 12-04-1995 10-11-1994 21-09-1995 07-07-1998
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WO 9625920 A	29-08-1996	US 5744125 A AU 702553 B AU 4926596 A BR 9607287 A EP 0820275 A JP 11501341 T	28-04-1998 25-02-1999 11-09-1996 23-06-1998 28-01-1998 02-02-1999
JP 03077813 A	03-04-1991	JP 2823891 B	11-11-1998
US 5122461 A	16-06-1992	DE 4137944 A JP 4311394 A	21-05-1992 04-11-1992
WO 9200373 A	09-01-1992	AU 695044 B AU 3010695 A AU 8295491 A CA 2086417 A CA 2166818 A EP 0547065 A JP 6500011 T KR 149181 B US 5837505 A US 5814495 A US 5631151 A	06-08-1998 22-02-1996 23-01-1992 30-12-1991 30-12-1991 23-06-1993 06-01-1994 17-08-1998 17-11-1998 29-09-1998 20-05-1997
EP 0363792 A	18-04-1990	AT 117376 T AU 4418989 A CA 2000114 A,C DE 68920685 D DE 68920685 T ES 2069560 T KR 9405597 B WO 9004029 A US 5837505 A US 5814495 A US 5631151 A	15-02-1995 01-05-1990 03-04-1990 02-03-1995 24-05-1995 16-05-1995 21-06-1994 19-04-1990 17-11-1998 29-09-1998 20-05-1997

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PC 00/01276

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0363792 A		ZA 8907501 A	27-06-1990
JP 57014517 A	25-01-1982	NONE	
WO 9323480 A	25-11-1993	IT 1255146 B	20-10-1995
		AT 161872 T	15-01-1998
		AU 4020593 A	13-12-1993
		DE 69316206 D	12-02-1998
		DE 69316206 T	23-07-1998
		EP 0639209 A	22-02-1995
		JP 8500371 T	16-01-1996
WO 9412644 A	09-06-1994	US 5340734 A	23-08-1994
		AU 689093 B	26-03-1998
		AU 5896594 A	22-06-1994
		CA 2149764 A	09-06-1994
		EP 0672148 A	20-09-1995
		JP 8503613 T	23-04-1996
		US 5466592 A	14-11-1995
		US 5792649 A	11-08-1998
		US 5486351 A	23-01-1996
		US 5801047 A	01-09-1998
		ZA 9308581 A	21-02-1995

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
15 March 2001 (15.03.2001)

PCT

(10) International Publication Number
WO 01/18125 A1

(51) International Patent Classification⁷: C09B 61/00,
69/10, A61K 7/42

(21) International Application Number: PCT/IB00/01276

(22) International Filing Date:
8 September 2000 (08.09.2000)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
MI99A001896 9 September 1999 (09.09.1999) IT

(71) Applicant and

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(BR).

(81) Designated States (national): AE, AG, AL, AM, AT, AU,
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ,

DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR,
HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,
LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ,
NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM,
TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM,
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian
patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European
patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,
IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG,
CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

- With international search report.
- Before the expiration of the time limit for amending the
claims and to be republished in the event of receipt of
amendments.

For two-letter codes and other abbreviations, refer to the "Guid-
ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.

(54) Title: SYNTHETIC VEGETAL MELANINS, PROCESS FOR THEIR PRODUCTION AND COMPOSITIONS CONTAIN-
ING THEREOF

(57) Abstract: The present invention relates to synthetic vegetal melanins obtained by polymerization of plant polyphenols, alone or
in combination with the eumelanin precursors. Chemical and enzymatic methods for production of said synthetic vegetal melanins
are also claimed, as well as their use for the manufacturing of cosmetic and pharmaceutical compositions.

WO 01/18125 A1

SYNTHETIC VEGETAL MELANINS, PROCESS FOR THEIR PRODUCTION AND COMPOSITIONS CONTAINING THEREOF

FIELD OF THE INVENTION

The present invention relates to high molecular weight synthetic vegetal melanins obtainable by polymerization of plant polyphenols.

More precisely, the present invention relates to synthetic vegetal melanins characterized by reproducible physical features, hereby obtainable by a controlled polymerization of a known amount of purified plant polyphenols, alone or in combination or in conjunction with eumelanin precursors.

The invention further relates to processes for the manufacture of said synthetic vegetal melanins either by chemical or enzymatic synthesis.

The invention further relates to the use of said synthetic vegetal melanins in cosmetics for the protection of the skin, of the mucous membranes and of hair against damages due to UV exposure and to oxidative environment, as well as their use for the manufacturing of pharmaceutical preparations and nutritional supplements exerting anti-inflammatory and immunomodulation activities.

BACKGROUND OF THE INVENTION

Melanins are complex three-dimensional biopolymers with several biological functions including: photoreceptor shielding, thermoregulation, photoprotection, camouflage, metal chelation and free radical scavenging.

The term melanin refers to a variety of highly irregular heteropolymers containing several structural moieties, such as free carboxy, amino, hydroxy, carbonyl, methoxy, biphenolic, indolic, quinoic groups and so on.

A broad classification of these photoabsorbing biopolymers can be made on the basis of the starting monomers (the actual melanin precursors), major classes being: eumelanins, pheomelanins and allomelanins.

Eumelanins are brown-black nitrogenous pigments produced by the rearrangement and oxidation of L-dopa and its metabolites. A type of eumelanins are the "oxymelanins" exhibiting tetrahydroxy indoles units within their polymeric structure; other eumelanins are the "neuromelanin" from dopamine (which is produced L-dopa carboxylase) and its metabolite, 3,4-dihydroxycarboxylic acid, as well as by neural

catecholamines and tetrahydroxyisoquinolines, e.g. occurring in the brain *substantia nigra*.

Pheomelanins are yellow-reddish pigments containing both nitrogen and sulfur atoms, which arise from the rearrangement and polymerization of cysteinyl-dopa, the condensation product of L-dopa and glutathione with further enzymatic hydrolysis. The pheomelanins generally result from the oxidative condensation of 1,4-benzothiazines; further pheomelanins are the "trichocromes" containing a bi-benzothiazinic chromophore ($-S-C=C-C=N-$); as well as condensed 1,4-benzothiazinalanines (e.g. in tricochrome C).

Allomelanins are highly colored pigments typically occurring in the plant kingdom, which may be further classified as "vegetal melanins" (alias "phytomelanins"), when resulting from the polymerization of plant polyphenols; and "fungi melanins" (alias "micomelanins"), such those containing naphthoic groups.

All melanins originate in nature from the common step of the conversion from monophenolic substances into the related ortho-diphenolic moiety. Noteworthy, this starting point of melanogenesis is always under strict enzymatic control.

Within the plant kingdom, the hydroxylating enzymes convert the phenolic precursors in ortho-diphenols: typically L-dopa to L-tyrosine occur in *Vicia faba* hull; p-hydroxybenzoic acid (PHBA) to protocatechuic acid in wheat seed hull; tyrosol to hydroxytyrosol in olive peel; p-coumaric acid to caffeic acid in green coffee skin; apigenin to luteolin in citrus peel; kaempferol to quercetin in clover blossom; pelargonin to cyanidin in grape fruit; genistein to glycitein in soybean hull, as depicted in FIG. 1 attached.

Diphenols undergo fast oxidation in presence of oxygen to provide semiquinones and quinones, which randomly polymerize and cross-link.

Some authors claim that some polymerization steps in melanogenesis may be under enzymatic control, e.g. the cyclization to indoles or decarboxylation, but the fact is quite controversial. As an example, eumelanins may be always produced by chemical synthesis, whereas the tyrosinase enzyme seems to be unable to produce eumelanin from the preformed indolic eumelanin intermediates.

The enzymatic hydroxylation of the monophenolic substrates is particularly intense

during ripening or post-ripening browning of fruits and vegetables, and further enhanced under specific stress conditions such as by handling, chopping, cutting, which increase vacuoli breackages, and consequently the release of the enzymes which prompts the formation of vegetal melanins. As in the plant kingdom the polymerization occurs extracellularly, enzymes should not contribute to the final steps of the melanin polymer formation.

Melanins found increasing acceptance as ingredient in health care compositions thanks to their photoprotective, anti-ageing, radical scavenging, chelating and tanning properties.

However, vegetal melanins are not currently available for industrial applications, and industrial cosmetic and pharmaceutical applications are limited to the use of the commercially available brown-black synthetic eumelanins or sepiomelanin, a natural eumelanin which is recovered from squid or octopus inks through a complex purification process.

On the other side, there were evidences that the vegetal melanins extracted from plants may provide an excellent dermoprotection, with anti-inflammatory activity and immunomodulation, by the inhibition of the effects on paw oedema and adjuvant induced disease, as described by Avramidis N. et al. (*Arzneimittelforschung*, 48(7):764-71, 1998).

The synthesis of novel pseudo-melanins is described in WO96/25290. However, these are fully artificial melanins, as there are no evidences of melanins synthesized in nature from precursors such those described in the aforementioned patent, i.e. from aloin and related anthrones or from p-diphenols and/or hydroxyanilines.

The first oxidation steps of caffeic acid was studied by John M; Gumbinger HG and Winterhoff H *Planta Med*, 56(1):14-8 (1990); whilst polyphenoloxidase in relation to browning on fruits was studied by Macheix J.J. et al., in *Crit. Rev. Food Sci. Nutr.*, 13(4):441-86 (1991) and Prabha T.N. et al., in *Ind. J Bioch. Biophys.*, 13(6):402-5 (1981).

The extraction of vegetal melanins is disclosed in WO00/09616, wherein melanin polymers are obtained by extraction procedures from suitable plant material. However, the extraction route consists in an expensive and time-consuming multi-

step procedure, yielding small amounts of vegetal melanins, so that the product is not rendered available for large scale application.

Moreover, the extraction methods provide only the oligomers of vegetal melanins, as high molecular weight melanins are not extracted, due to their strong bounding within the vegetal matrix formed by the other vegetal biopolymers, and because of their low solubility in common solvents.

Finally, the extracted vegetal melanin may suffer a broad range of variability in their physico-chemical properties, due to the large variation of the both polymerization condition and the plant polyphenols composition, for example caused by the climatic variation along the year, the different cultivars, and the different grade of ripening.

BRIEF DESCRIPTION OF THE DRAWINGS

Some illustrative embodiments of the invention and examples of carrying out the invention, are described in detail hereinbelow, with reference to the accompanying drawings in which:

FIG. 1 is schematic diagram of the conversion of some representative plant monophenols into diphenols, the actual vegetal melanin monomers;

FIG. 2 is a schematic flow diagram illustrating the Raper-Mason pathway from main eumelanin precursors, whose condensing centers are highlighted by double arrows;

FIG. 3 to 6 are schematic view of UV-visible spectra of synthetic vegetal melanins according to the present invention.

DETAILED DESCRIPTION OF THE INVENTION

It is commonly understood that products intended for use in industrial preparations must show constant parameters, be reproducible and free from contaminants.

We have now found out that synthetic vegetal melanins showing reproducible physical features may be conveniently produced in vitro.

We have also found out that high molecular weight synthetic vegetal melanins showing reproducible physical characteristics may be conveniently produced in vitro.

In fact, we have ascertain that in in vitro oxidative conditions, the ortho-diphenolic moiety of vegetal melanins precursors, such as the polyphenols are converted into semiquinones and quinones, thus making the diphenolic ring available to polymerize,

either by self-condensation or copolymerization with other polyphenols and/or with eumelanin precursors.

We have ascertain that high molecular weight synthetic vegetal melanins having reproducible features may be produced starting from a given composition of suitable monomers submitted to controlled polymerization conditions.

Therefore, according to one of its aspects, the present invention concerns synthetic vegetal melanins, more specifically the synthetic vegetal melanins herein defined as omo- and ethero-polymers, obtained by in vitro polymerization of purified plant polyphenols, optionally with copolymers such as the eumelanin precursors, more particularly obtained by in vitro polymerization of at least one monomer unit (a), and optionally at least one compound (b), wherein:

(a) is a plant polyphenol; and

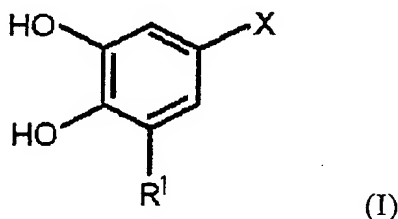
(b) is an eumelanin precursors;

said synthetic vegetal melanin being characterized by a Red:Green:Blue (RGB) ratio comprised between 1 : 0.88 : 0.84 and 1 : 0.64 : 0.41.

Preferably the molecular weight of at least 80% of the synthetic vegetal melanins of the present invention is $\geq 0.8 \times 10^4$ dalton.

The synthetic vegetal melanins of the present invention are characterized by reproducible physical characteristics, such as color, UV-visible spectra, and the antioxidant activity.

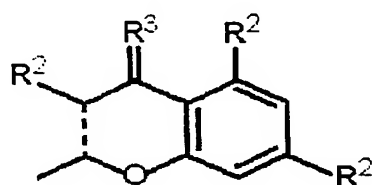
According to a preferred embodiment, the invention concerns synthetic vegetal melanins obtainable by polymerization of at least one plant polyphenol of formula (I):



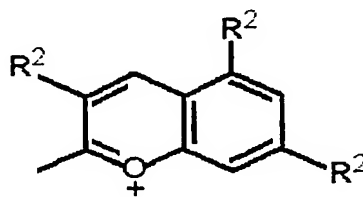
wherein:

R^1 represents hydrogen, hydroxy, or methoxy group; and

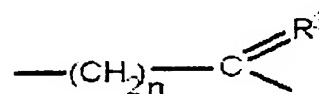
X represents a residue of formulas (II), (III) or (IV)



(II)



(III)



(IV)

wherein:

n is 0, 1 or 2;

R² represent, each independently, hydrogen, hydroxy, methoxy, or an O-glycosilated group;

R³ represents, one oxygen, two hydrogen atoms, or one hydrogen and one hydroxy group; and
the phantom line between C₂ -C₃ represents a double or a single bond,
as well as salts, esters and solvates thereof.

In controlled oxidative conditions, the plant polyphenols of formula (I) are converted into the corresponding semiquinones and quinones, which afterward undergo either self-condensation or copolymerization with other plant polyphenols and/or with eumelanin precursors.

The synthetic vegetal melanins of the present invention, are thus primarily characterized by distinctive and reproducible physico-chemical features, such as the molecular weight, the color and their spectroscopic features, thanks to the control of the starting monomeric composition and of the polymerization conditions.

The main distinctive feature of the synthetic vegetal melanins are their color. In fact, all melanins exhibit a broad band of absorption within the UV-visible range, with a smooth decreasing pattern toward high wavelengths, with no detectable maxima. Therefore the UV-visible spectrum may not clearly identify any single melanin, whereas the color tones are clearly distinguished by a color analyzer. The color space dimension is evaluated by a three-points parameter, such as the Red, Green, Blue (RGB) ternary value. Moreover, when the RGB ternary value is expressed as R:G:B ratio, it univocally identify the melanin by its color

characteristic, hence regardless of the concentration of the sample.

It is well known that the color intensity greatly influence the color perception, therefore a color definition will greatly differ according to the sample concentration. On the other side the R:G:B ratio of the synthetic vegetal melanins of the present invention is a costant and reproducible feature. Moreover, it was surprisingly found that the R:G:B ratio does not generally differs when the melanin is in ionic (salt) form, such as the sodium salt, or in non-ionic (acidic) form.

In that respect they clearly differ from the natural occurring vegetal melanins, which may vary in a broad range of chemical composition and/or molecular weight due to the seasonal and specimen variation of the concentration and composition of the plant polyphenols, as well as the polymerization conditions such as temperature, humidity, oxygen partial pressure on the cultivated land and so on.

The synthetic vegetal melanins of the present invention are thus obtained by an oxidation process, also called melanization process, starting from highly purified precursors in presence of atmospheric oxygen, to afford synthetic vegetal melanins of high molecular weight, substantially higher than 80%, as calculated using the elution times from a precalibrated molecular sieve HPLC column.

The synthetic vegetal melanins of the present invention, thus produced in presence of atmospheric oxygen from highly purified selected monomers, i.e. plant polyphenols and optionally eumelanin precursors, are characterized by distinctive photoabsorption patterns.

The in vitro ex-vivo antioxidant test carried out on synthetic vegetal melanins of the present invention exhibited reproducible antioxidant activities. Generally the inhibition of the metal-dependent oxidation (secondary oxidation) is higher than the radical scavanging activity (primary oxidation) in model system. This is due to the capacity to form polydentate complexes with the pro-oxidant metals, with significant inhibition of Fenton-like reactions.

The synthetic vegetal melanins of the present invention are further characterized by a large availability of colors, thus are particularly suitable for a

variety of application in the health care industry.

The term "plant polyphenol " as used in the present invention designates those molecules bearing an ortho-dihydroxyphenolic moiety typically occurring as plant secondary metabolites.

5 Preferred plant polyphenols are those polyphenols of formula (I) above.

Common plant polyphenols may either have a polycyclic structure, such as flavonoids, or an open-ring diphenolic structure.

10 Illustrative example of flavonoids are flavonols such as quercetin and its glycosides, as the 3-rhamnoside (quercetrin) from many plants (e.g. horse chestnut) or the 3-rutinoside (rutin) from grape, tobacco and eucalyptus plants; flavons such as fisetin and fustin from Venice sumac; or mixture of flavonoids and benzophenones such as in morin and maclurin from old fustic (*Morus tinctoria*).

15 Further illustrative example of flavonoids are those contained in grape and in the peel of other edible fruits, thus containing flavanols such as myricetin, flavones such as luteolin, flavandiols such as dihydroquercetin, as well as the flavanols. Grape contains also the anthocyanins and chalcones, which are the biosynthetic precursors of flavonoids and anthocyanins, the latter being slowly
20 reverted to the corresponding chalcones when kept in neutral or slightly alkaline aqueous solution. Anthocyanins are the coloured matter of flowers and also occurring in coloured fruits and vegetables, including cyanidin and delphinidin, which may also be substituted as mono, di-saccharides, and position 3 may be acylated, e.g. with p-coumaric acid.

25 Further illustrative example of flavonoids are the oligomeric proanthocyanidins (OPC), precondensed oligomer of flavanols, i.e. (+)-catechin and (-)-epicatechin, which are commonly extracted from grape seed, pine bark, cocoa and other vegetal sources rich in flavan-3-ols in the free, glucosylated, esterified, or condensed forms. Galloylated catechin monomers are typically extracted from green tea or Catechu gambir, which contain (+)-gallocatechin (GC), (+)-gallocatechin gallate (GCG), (-)-epigallocatechin (EGC) and (-)-epigallocatechin gallate (EGCG).

30 Further polyphenols for our purposes may include those plant diphenols which are biogenetically correlated with flavonoid, e.g. the

benzyndenpyranones such as brazilin from brazilwood and sappanwood, and haematoxylin from logwood; or the isoflavones such as glycitein from soybean.

Other plant polyphenols which may be used in the present invention are the open ring dihydroxyphenols.

5 Illustrative example of open ring dihydroxyphenols are hydroxytyrosol and its esters (oleuropein, verbascoside, etc.) found in olive fruit and leaves; protocatechuic acid found in most higher woody plants and in wheat; and protocatechuic aldehyde, found in vanilla, and other phytoalexins.

Further illustrative example of open ring dihydroxyphenol are gallic
10 acid and its derivatives, which may be obtained from hydrolisis of plants rich in tannins, such as nutgalls. Tannins (alias tannic acid) are also classified as ellagitannins or gallotannins, the latter releasing gallic acid by hydrolisis treatment.

Plant polyphenols suitable for our purpose may occur either in the free form (aglycones) or as natural occurring esters and ethers (e.g. glycosides, gallates,
15 and the like) as well as in oligomeric form (e.g. proanthocyanidins and viniferins). Glycosides are the O-derivatives of the plant polyphenols in the plain form (aglycones), generally substituted in one or two hydroxy groups in the 3, 5 or 7 positions with naturally occurring mono- di- or trisaccarides, such as ruthoside, galactoside, rhamnoside, glucoside, sophoroside, rhamnoglucoside, and the like.

20 Plant polyphenols suitable for the present invention may be in form of titolated plant extracts with known chemical composition, such as obtained by standardized extraction methods from vegetal sources, either in solid form, e.g. dry extracts, or in liquid form, e.g. hydroalcoholic or as an alkaline solution.

In the present invention, plant polyphenols are purified compound,
25 which are in the form of enriched fractions or even as pure compounds, either purified from natural sources or produced by chemical synthesis, so as to achieve synthetic vegetal melanins having the desired compositions, their physical features being therefore industrially reproducible. Another important technical advantage of using plant polyphenols as enriched fractions or even as pure compounds is that
30 formation of foam during the polymerization reaction is thus reduced.

In a further embodiment of the present invention there are provided

"mixed-type" synthetic vegetal melanins, thus produced by the oxidative copolymerization of plant polyphenols with eumelanin precursors.

As shown in FIG. 2 by the Raper-Mason pathway (Physiol. Rev. 8. 245, 1928; J. Biol. Chem. 172, 83, 1948) the typical eumelanin precursors are L-dopa and
5 its melagenetic intermediates: dopa-semiquinone, dopaquinone, leucodopachrome, 5,6-dihydroxyindole-2-carboxylic acid (DHICA), dopachrome, 5,6-dihydroxyindole (DHI), and indole-5,6-quinone.

L-dopa occurs in several plants, such as *Mucuna pruriens* and *Vicia Faba*; DHICA and DHI occurring in plants such as *Rhaphidophora korthalsii*; 3,4-dihydroxyphenylethylamine (dopamine) is occurring in vegetables and fruits, such as
10 in the peel of banana or potatoes; pyrocatechol present in some vegetable such as garlic; and pyrogallol found in geranium and *Hypericum p.*

Preferred eumelanin precursor for the present invention are dopamine and L-dopa, both being partially polymerized "as such" and partially in situ converted into typical
15 eumelanin indolic precursors by contact with oxygen, as per Raper-Mason pathway.

The synthetic vegetal melanins of the present invention can be prepared from plant polyphenols, with or without eumelanin precursors, by a number of chemical or enzymatic polymerization procedures.

A preferred method of producing synthetic vegetal melanins is the
20 chemical method, i.e. bubbling air or oxygen through an alkaline aqueous solution of the monomers at pH 10 or above (e.g. by 1 N sodium hydroxide or 2-3 N ammonia) during 12 to 48 hours (preferably 24 hours), at a temperature ranging from 10 to 90°C, preferably in the range of temperature from 20 to 30°C. The alkaline aqueous solution may comprise catalytic amounts of pro-oxidant metals, such as Cu^{++} , Fe^{++} ,
25 Ni^{++} , or Co^{++} solutions, for example at 5 to 20 mM concentration.

The amount of alkali depends on the chosen plant polyphenol, i.e. the strength and quantity of ionizable hydrogen atoms to be salified in order to achieve the full solubilization of the starting monomers. For strong alkali such as NaOH or KOH the optimum ratio with the melanin precursors is around 3:1 equivalents,
30 whereas for weak alkali such as ammonia the optimum ratio with is around 10:1.

A fast polymerization procedure may also be carried out by using

chemical oxidizing agents, such as hydrogen peroxide, hydrogen iodide, ammonium persulfate, potassium permanganate, or magnesium perchlorate. If such strong oxidants are employed, the synthetic vegetal melanin precursors may also be present as monohydroxylated (phenolic) form, and thereafter converted in situ to the corresponding diphenol compounds (the actual melanin monomers) in a "one-pot" oxidative polymerization reaction.

A further preferred method of producing synthetic vegetal melanins of the present invention is the enzymatic synthesis by suitable melanin-forming enzymes, such as tyrosinases, polyphenoloxidases (catechol oxidases), phenolases, (phenoloxidases), peroxidases, laccases, lipoxygenase, and mixture thereof.

The enzymatic process makes use of a stream of air or oxygen through a buffered aqueous solution of plant polyphenols and/or eumelanin precursors during 12 to 48 hours or more, preferably at controlled temperature in presence of a suitable enzyme. In this case, the synthetic vegetal melanin will be produced in low concentration, due to their low solubility in water of the plant polyphenols, present in their neutral (non-ionic) form.

Suitable enzymes for the synthesis of the synthetic vegetal melanin of the present invention may be obtained from several melanin-producing biological cells or tissues, such as:

- (1) extracts of animal skins, e.g. horses, cattle, sheep, pigs, or any other such mammalian source; extracts of skins of fish, amphibia, reptiles, birds;
 - (2) extracts of melanin-producing vegetal source, e.g. mushrooms, potatoes, bananas;
 - (3) extracts of melanin-producing invertebrate organisms, e.g. worms, arthropods;
 - (4) extracts of melanin producing microorganism, e.g. bacteria and protozoans;
 - (5) extracts of any organisms in which genes for enzymes producing soluble melanin have been genetically cloned; and
 - (6) culture media of any organisms or cells which secrete enzymes for producing soluble melanin-such organisms or cells may or may not express cloned genes for the enzymes.
- Some of the above mentioned enzymes are commercially available and the others can be prepared by known techniques.

Furthermore, the enzymes capable of actively synthesizing melanin may not necessarily have to be isolated from the organism producing thereof.

Particularly when the enzyme applied retaining a significant hydroxylating activity, the monomers of synthetic vegetal melanin may be formed in situ from the
5 corresponding monophenols (the "pre-monomers"), as in the case of the chemical synthesis in presence of chemical oxidizing agents.

Illustrative examples of pre-monomers of synthetic vegetal melanins include dihydrokaempferol, armadendrin, apigenin, p-hydroxybenzaldehyde, PHBA, tyrosol, p-coumaric acid, kaempferol, pelargonin, and genistein.

10 Illustrative examples of pre-comonomers of synthetic vegetal melanins include L-tyrosine, tyramine, and 5-hydroxy-indole.

The polymerization may be carried out in a standard fashion, i.e. by introducing all the monomers at once, as well as by separate or sequential polymerization of a portion of monomers, as to provide melanin "block co-
15 polymers".

The synthetic vegetal melanins may be used for their application either as the native solution or in solid form, the latter obtained by a number of precipitation procedures known to those skilled in the art, for example by addition of an organic or mineral acid, e.g. HCl, H₂SO₄, acetic acid, glycolic acid, or L-tartaric
20 acid; or by addition of water-soluble organic solvents, for example 2 volumes of acetone or ethanol; or by addition of divalent or trivalent metal salts, for example solution of ZnCl₂, MgCl₂, AlCl₃, or CaCl₂, thus forming insoluble metal complexes; or by mixed techniques.

Moreover, the synthetic vegetal melanins may be stabilized by stopping the
25 polymerization by the addition of boric acid or salt thereof, which are added to the final reaction mixture to prevent further increase of the molecular weight.

Further modification to the structure of the synthetic vegetal melanin may be applied in order to alter their chemical and physical properties.

Esters of synthetic vegetal melanins can be prepared by standard
30 methods of acylation, such as by the reaction of the melanin with acid chlorides or acid anhydrides in the presence of base. Of particular utility is the preparation by

reaction with acid anhydrides in pyridine as utilized for the acylation of certain other lower molecular weight flavonoids by Thompson, et al., JCS, 1387 (1972), Fletcher, et al., JCS, 1628 (1977) and Hemingway, JCS, 1299 (1982).

Ethers can be prepared by standard methods of etherification such as
5 by reaction of the synthetic vegetal melanin with alkyl halides and alkyl tosylates in bases. Methyl ethers are readily prepared by the use of diazomethane as utilized in the etherification of certain other low molecular weight flavonoids by Thompson, JCS, 1387 (1972), Hemingway, JCS, 1299 (1982).

The synthetic vegetal melanins may be isolated and/or purified by
10 filtration, ultrafiltration, fractional sedimentation, freeze-drying, spray-drying, dialysis, centrifugation or combination thereof.

Alternatively, the synthetic vegetal melanins of the invention may be directly used as aqueous solutions, then complexed or compounded with suitable ingredients during the preparation of cosmetic or pharmaceutical compositions. When the synthetic
15 vegetal melanins are used in a soluble or suspended form in a fluid preparation, they are preferably associated with at least one part in weight of a suitable surfactant, such as cationic, anionic, amphoteric and non-ionic surfactants, in order facilitate the dispersion and stabilize the biopolymer during storage.

The synthetic vegetal melanins of the present invention may be used
20 in pharmaceutical and cosmetic preparations to provide protection against oxidative stress, due to the presence of the highly active vegetal moiety within the polymeric network.

In addition, the synthetic vegetal melanins of the present invention can be produced in a large variety of colors, thus being a valuable tool for producing
25 cosmetic compositions for facial make-up and hair dyes, as well as in tanning, anti-sun, moisturizing-protecting compositions and toiletry preparations.

It is a further object of the present invention to provide cosmetic compositions comprising the aforementioned synthetic vegetal melanins.
The cosmetic composition according to the invention may further comprise any
30 cosmetically acceptable ingredients, for example those included in the INCI list drawn by the European Cosmetic Toiletry and Perfumery Association (COLIPA) and

issued in 96/335/EC "Annex to Commission Decision of 8 May 1996" and further modifications.

Preferred cosmetic ingredient to be associated at the synthetic vegetal melanins in fluid cosmetic formulations are the surfactants, thus including anionic, cationic, nonionic and zwitterionic surfactants, either with medium-low molecular weight or as oligopolymer and polymer containing a polarized moiety within a predominantly hydrophobic structure.

Among surfactants are particularly preferred those bearing a positive charge, thus capable of forming a ion pair with the substantially negatively charged synthetic vegetal melanins, therefore stabilized and kept in a suspended/solubilized form.

More preferably, the synthetic vegetal melanins in solution or in a precipitated form are premixed with one or more surfactant (e.g. soybean lecithin and mono- diglycerides) prior the mixing with the rest of the formulation, to produce a stable color, an homogeneous appearance and enhancing stability and shelf life of the cosmetic formulations containing thereof.

The cosmetic composition of the invention can be formulated as a lotion, a fluid cream, a cream or a gel. The composition can be packaged in a suitable container to suit its viscosity and intended use by the consumer. For example, a lotion or fluid cream can be packaged in a bottle or a roll-ball applicator, or a capsule, or a propellant-driven aerosol device or a container fitted with a pump suitable for finger operation. When the composition is a cream, it can simply be stored in a non-deformable bottle or squeeze container, such as a tube or a lidded jar.

The invention also refers to pharmaceutical and/or nutritional compositions comprising synthetic vegetal melanins, which may provide anti-inflammatory and immunomodulation activities by the oral intake. The synthetic vegetal melanins may be also used in oral compositions for their coloring properties, further associated with an antioxidant behaviour to increase the shelf-life of the edible products comprising thereof.

The oral compositions comprising synthetic vegetal melanins to be administered by the oral route in solid form such as sachets or soft gelatin capsules, fast or slow-release tablets, or in liquid or emulsified forms, and may also include further active

ingredients and acceptable excipients, for examples those described in "Remington's Pharmaceutical Sciences Handbook", Mack Pub. Co, USA.

According to another of its aspects, the invention relates to a method for treating and/or preventing diseases due to exposure to radiations or to oxidative environment as well as in inflammatory diseases or immunitary unbalance, which comprises administering to a subject an effective amount of at least a synthetic vegetal melanin of the invention.

Furthermore, the synthetic vegetal melanins may be used as food ingredient (additive) for the manufacturing of health food, dairy products and canned food, as well as in beverages such as soft drinks, syrups and fruit juice.

The following examples are preferred features of the invention but are not intended to limit it in any way.

EXAMPLES

Method A – Chemical synthesis of synthetic vegetal melanins

A weighted amount of a total monomers, calculated from the actual polyphenolic content of the starting material, is charged into a flat-bottom conical flask (500 ml) and solubilized in 200 ml of sodium hydroxide 1 N (the amount of sodium hydroxyde shall undergo minor variation according to the K_a and the number of acid centers within the molecule of each plant polyphenol, i.e. according to the quantity and quality of ionizable hydrogen atom to be salified in order to achieve the full solubilization of the starting monomers).

The flask, placed in a thermostatic bath at 24°C, is equipped with an air pump (Silent Air[®], manufactured by Renn-Plax, Inc. in Taiwan R.O.C.) connected by a flexible hose to a disperser, which is placed at the bottom of the flask. The air is forced through the disperser and finely divided throughout the alkaline solution, which, for the oxidative polymerization, is kept under air bubbling for a time ranging from 12 to 48 hours at a temperature of 24°C. The water lost due to the evaporation caused by the stream of air was refilled from time to time in the reaction vessel.

The reaction mixture readily develops colors, as vegetal melanins absorb widely throughout the ultraviolet and visible spectra (e.g., 220 to 700 millimicrons), the final color of the melanin solution varying according to the monomer composition and, at

less extent, to the reaction time.

Absorbance of vegetal melanins was measured by spectrophotometric absorbance of a solution obtained by dissolving 4 mg of synthetic vegetal melanin in 100 ml of 0.1 N NaOH. The UV visible absorption spectrum was recorded on a spectrophotometer
5 between the wavelength of 200-700 nm in 1 cm path length cuvette. The representative spectra (200-500 nm) of a selected number of vegetal melanins are shown in FIG 3, 4.

The aforementioned procedure were modified to provide further vegetal melanins, which are modified from the original omo- or etheropolymer under the circumstances
10 described herewithafter.

Modification applicable to the method A

i) The use of pro-oxidant transition metal ions, such as Cu^{++} , Ni^{++} , Co^{++} , Fe^{++} to enhance oxidative polymerization.

When pro-oxidant metal ions are added to the reaction mixture, a darker tone is
15 generally obtained, the effect being low for vegetal melanins comprised of flavonoid-type plant polyphenols, higher in the presence of plant polyphenols with open catechic structure and even more pronounced with mixed-type vegetal melanin, e.g. with high L-Dopa to plant polyphenols ratio.

In the next Examples, 1,25 mM Cu^{++} solution is formed by solubilizing 20 mg
20 $\text{CuSO}_4 \cdot 2\text{H}_2\text{O}$ into 100 ml of the reaction mixture.

ii) The use of weak bases instead of NaOH or KOH solutions.

The vegetal melanins display a generally darker tone in presence of ammonia or other ammonium bases (e.g. triethanolamine) as alkalifying agent. Noteworthy, in ammonia solution there is low, if any, catalytic activity of pro-oxidant metals salt
25 such as Cu^{++} , which shall be due to the partial inactivation by the Cu-ammonium complex.

In the next Examples, ammonia 2.5 N is used as alkalifying medium instead of sodium hydroxide.

iii) The use of chemical oxidant, organic or inorganic peroxides.

30 The strong oxidating agents, such as persulfates, KMnO_4 or hydrogen peroxide, may be added to the reaction mixture to produce synthetic vegetal melanins. Moreover,

these chemicals are capable of converting the pre-monomers in the actual melanin precursors, so that the corresponding monophenols can be used instead of the diphenolic monomers, and converted in situ to the actual precursors by the oxidant agent, finally being polymerized by the combined activity of ambient oxygen and chemical oxidation.

If 40 ml/l of H_2O_2 20 vol is added dropwise to the polymerization mixture during the reaction additionally a pale color is generally obtained. The UV spectrum of bleached vegetal melanins shows a absorption flattening at higher wavelength. The bleaching effect is comparatively higher on vegetal melanin with low plant polyphenols to eumelanin precursor ratio.

In the next Examples, ammonium persulfate in 2:1 molar ratio as regard to the total pre-monomer content is added to the reaction mixture under high stirring to provide oxygen circulation, for 24 hours. At the end of reaction a brown-red vegetal melanin is obtained.

Method B1 – Enzymatic synthesis of the vegetal melanins by laccase

A weighed amount of a total plant polyphenols, calculated from the actual monomer content of the starting material, is charged into a flat-bottom conical flask (250 ml) and solubilized in 100 ml of a solution 20 mg of laccase at 0.8 LAMU/mg (Denilase[®] II S, Novo Nordisk A/S, Bagsvaerd, Denmark) in phosphate buffer 50 mM at pH 6.5.

The solution is kept at a temperature of 37°C under high stirring by a magnetic bar, so to force hair circulation for 24 hours. At the end of reaction insoluble, colored vegetal melanins are obtained.

Method B2 – Enzymatic synthesis of the vegetal melanins by tyrosinase

A weighed amount of a total plant polyphenols, calculated from the actual monomer content of the starting material, is charged into a flat-bottom conical flask (250 ml) and solubilized with 20 mg of mushroom tyrosinase at 2200 U/mg polyphenoloxidase (Sigma, St.Louis, USA) in 800 ml phosphate buffer at pH 6.5 under stirring. The solution is kept at a temperature of 24°C under high stirring to provide brown-black vegetal melanins.

Modification applicable to both method A and Bs

iv) The use of boric acid or its soluble salts to stop post-polymerization pathways.

When borate and melanins are contacted in solution, a bidentate link of the orthoboric structure and the ortho-quinoid moiety of the biopolymer to provide a cyclic ester is postulated to occur, thus preventing the further darkening and post-polymerization of the synthetic vegetal melanins.

- 5 In the next Examples, boric acid is added 30 minutes before the end of the reaction at about 1:2 molar ratio with respect to the total amount of the starting vegetal monomer units.

v) The use of anti-foaming agent during oxidative polymerization.

- The use of anti-foaming agents at low concentration is needed to avoid excess foam formation during water injection for plant polyphenols from medium purity extracts, thus with a residual content of proteins, saponins and other naturally occurring, foam-making substances.

In the next Examples, Dow Corning[®] 2210 is added at 5-40 ppm into the reaction mixture.

- 15 C) Precipitation of the synthetic vegetal melanins

The vegetal melanins may be obtained in solid form after precipitation by the addition of acid, of water-soluble organic solvents, or solution of di- or trivalent metal salts, and combination thereof.

- In the next Examples, precipitation of vegetal melanins was carried out by a addition of one equivalent ZnCl_2 1M in water, then the reaction mixture was heated at 60°C for 5 minutes and cooled to ambient conditions. The slurry is washed with demineralized water, filtrated, then dried in nitrogen stream, and the dry solid is grinded.

Materials – Monomers and precursors of the synthetic vegetal melanins

- 25 - Quercetin $2\text{H}_2\text{O}$; MW = 338; content 99%; from Fluka Chemie AG (Buchs, CH).
- Rutin $3\text{H}_2\text{O}$; MW = 664.5; content 96%; from Austin Ch. Co. (Buffalo Grove, IL, USA).
- Anthocyanins with grape flavonoids (AGF) (4:1 mol/mol), average MW = 338.7 (calculated as delphinidin); 20% solution; source grape pomace; from Enocianina
30 Fornaciari Srl (Reggio Emilia, Italy).
- Grape seed oligomeric proanthocyanidins (GS-OPC); average MW = 290

(calculated as catechin monomer); content $\geq 90\%$; from Centroflora Ltd (Sao Paulo, Brazil).

- Green tea polyphenols (GTP); average MW = 316 (calculated as gallocatechin); content 95%; from Xinguang Ind. Products (Chengdu, Sichuan, P.R. of China).

5 - Fustic extract, 40% solution with morin and myricetin, average MW=302 (calculated as morin); from CE Roeper GmbH (Hamburg, Germany).

- Gallic acid H₂O; MW = 188; content $\geq 98\%$; from AlfaChem (N. C. Islip, NY, USA).

10 - Protocatechuic acid (PA); MW = 154; content 99%; from Ubichem Plc (Eastleigh, UK).

- Protocatechaldehyde (PAI); MW = 138; content 99%; from Ubichem Plc (Eastleigh, UK).

- Tannic acid; average MW = 634 (calculated as corilagin); content $\geq 90\%$; from Sigma-Aldrich Srl (Milan, Italy).

15 - Naringin; PM = 580.5; content 95%; from Freeman Industries Inc. (New York, NY, USA).

- L-Dopa; content $\geq 99\%$; MW = 197; content 99%; from Taiye Co. (Kunshan, P.R. of China).

20 - Dopamine; content $\geq 99\%$; MW = 153; content 99%; from Recordati Srl (Milan, Italy).

- Pytocatechol; content $\geq 99\%$; PM = 110; content 99%; from Borregard SpA (Ravenna, Italy).

- p-Hydroxybenzoic acid (PHBA); MW = 148; content 99%; from Fluka Chemie AG (Buchs, CH).

25 - p-Hydroxybenzaldehyde; MW = 122; content 95%; from Fluka Chemie AG (Buchs, CH).

- L-Tyrosin; content $\geq 99\%$; MW = 181; from Diamalt GmbH (Munich, Germany).

Examples 1-15 – Synthetic vegetal melanins

30 Following the standard procedure of Methods A and B1 or B2 and further modifications (from i to v), 100 ml of the solution of monomers of pure plant polyphenols, as shown in Table I, or of a mixed solution of monomers of pure plant

polyphenols and eumelanin precursors, as shown in Table II, were submitted to oxidative polymerization by dispersed air to provide synthetic vegetal melanins of different reproducible colors.

TABLE I

EXAM- PLE N°	SYNTHETIC VEGETAL MELANIN MONOMER (a)		SYNTHETIC VEGETAL MELANIN COMONOMER (b)		Molar ratio (a):(b) (mmol)	Reaction time (h)	NaOH 100 ml N	METHOD APPLIED (A, B, i-v)	Solid content (%)
	Plant polyphenol	(g)(mmol)	Plant polyphenol	(g)(mmol)					
1	quercetin	6	17,6			27	1,2	A	5%
2	quercetin	2,3	6,7			48	0,5	A	2,5%
3	gallic acid	5	26,3			48	1	A	5%
4	morin	27,78	36,8			24	1	A-v	10%
5	protocatechualdehyde	3,36	24,1		1:1	30	1	A-i	9%
6	rutin	5	7,2	protocatechuic acid	3,76	24	1	A-i,v	5%
7	hematoxylein	10	29,8			24	1	A	10%
8	GTP	4,5	13,4			24	1	A-v	10%
9	GTP	4,5	13,4			24	1/3 13a	A-iv	10%
10	OPC	3,5	5,4	gallic acid	2	36	NH3 2,5	A-ii	8%
11	anthocyanins	20	11,1	grape flavonoids	5	24	1	A	5%
12	OPC	4,167	6,5			24	1	A	4%
14	naringin	3,05	5,0			24	1	A-iii	10%
13	protocatechuic acid	0,075	0,5	protocatechualdehyde	0,069	24		BI	1%
14	GTP	0,170	0,5			24		BI	1%
15	anthocyanins	2	1,1			18		B2	1%

TABLE II

EXAM- PLE N°	SYNTHETIC VEGETAL MELANIN MONOMER (a)		SYNTHETIC VEGETAL MELANIN COMONOMER (b)		Molar ratio (a):(b) (mol:mol)	Reaction time (h)	NaOH 100 ml N	METHOD APPLIED (A, B, i-v)	Solid content (%)
	Plant polyphenol	(g)(mmol)	Plant polyphenol	(g)(mmol)					
16	AGF	18	L-dopa	6,48 32,6	1:3	30	0,9	A-v	10%
17	AGF	19,2	L-dopa	4 20,1	1:2	24	1	A	10%
18	protocatechuic acid	1,815	L-dopa	7,92 39,8	2:7	24	1	A	9%
19	quercetin	3	L-dopa	3 15,1	4:7	27	1,2	A	5%
20	quercetin	1,375	L-dopa	1,375 6,9	4:7	48	0,5	A	2,5%
21	gallic acid	5,61	L-dopa	6,6 33,2	8:9	24	1,1	A-i	10%
22	rutin	8	pyrocatechol	1,3 11,7	1:1	24	1	A-v	9%
23	protocatechualdehyde	7,2	dopamine	2,16 11,3	9:2	30	0,9	A-i	9%
24	GS-OPC	6,6	L-dopa	4,125 20,7	1:2	24	0,9	A	10%
25	GS-OPC	6,1	dopamine	3,65 19,1	1:2	24	0,8	A	10%
26	morin	22,22	L-dopa	2,389 21,5	4:3	24	1	A-v	5%
27	brazilein	3,75	dopamine	1,1 5,7	2:1	24	0,95	A-v	10%
28	brazilein	3,75	dopamine	1,1 5,7	2:1	24	1/2 15b	A-iv	10%
29	p-OH-benzoic acid	1,64	tyrosine	3,64 19,9	3:5	24	1	A-iii	10%
30	protocatechualdehyde	0,069	L-dopa	0,099 0,5	1:1	24		B1	1%
31	GTP	0,085	L-dopa	0,099 0,5	1:2	24		B1	1%
32	protocatechuic acid	0,065	tyrosine	0,182 1,0	1:2	24		B2	1%

Each Example was repeated at least twice in the same operative conditions, providing synthetic vegetal melanins which exhibit reproducible physical features, such as color, MW distribution and the UV-spectra, whose representative examples are shown in FIG. 3 to 6.

5 The color of the synthetic vegetal melanins of Example 1-32 were analyzed by according a standardized procedure based on scanning of a small quantity of sample adsorbed onto a white paper and the analysis of the color composition.

10 In brief, a quantity of 0.05-0.2 ml of the solution were deposited on a filter paper (Whatmann N.3), thereafter the spot were dried at room temperature. For the synthetic vegetal melanin obtained by oxidative polymerization in alkaline solution, a spot were treated with approximately 0.1 ml of HCl 1N to obtain the corresponding non-ionic form.

15 The spots were scanned with HP ScannJet 4300C[®] Pro (Hewlett Packard) set in a fixed mode for input, aquisition as 16.7 million colors photo, with an output set at customized resolution (x20), resolution area of 239 units, shadows at 6 units and medium tones at 2.2 units.

20 The spots were then analyzed in different point by the software WhatColor v3.21e by Hikaru Kakahara (Japan) with the dither scope set at 8 surrounding pixels for the analyze of the average color of the spot of vegetal melanins by chemical synthesis, where those by enzymatic synthesis where analyzed with a single pixel scope.

The value was given in decimal figure with the RGB composition of the 3 main colors, namely Red, Green and Blue, as shown in Table III and Table IV.

25 The R:G:B ratio are thus not depending on the concentration of the melanin and are thus useful for the evaluation of both reproducibly and for applicative purposes.

Example 1-32 were evaluated with the afore mentioned method to provide consistent color ranging from the more yellowish tones to the reddish or scarlet browns, with a number of neutral and graysh brown tones in the middle, as shown in Table III and Table IV.

TABLE III

EXAM- PLE N°	IONIC FORM (Na SALT) - RGB VALUES									NONIONIC FORM - RGB VALUES									R:G:B RATIO			AVERAGE R:G:B RATIO						
	MEDIUM			LIGHT SPOT			DARK SPOT			MEDIUM			LIGHT SPOT			DARK SPOT			IONIC FORM			NONIONIC			R:G:B RATIO			
	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	
1	122	98	72	136	109	79	111	91	68	195	149	109	238	206	164	163	121	89	1	0,81	0,59	1	0,80	0,61	1	0,80	0,60	
2	168	135	99	175	141	102	146	119	90	182	140	106	221	190	149	172	129	97	1	0,81	0,60	1	0,80	0,61	1	0,80	0,60	
4	138	119	94	147	127	99	130	113	90	196	167	133	208	179	143	177	148	117	1	0,87	0,68	1	0,85	0,68	1	0,86	0,68	
5	121	78	55	125	82	56	116	75	51	184	128	80	194	135	83	179	124	76	1	0,65	0,45	1	0,69	0,43	1	0,67	0,44	
6	68	64	63	71	65	64	55	54	54	116	97	87	125	104	94	111	89	79	1	0,94	0,93	1	0,82	0,74	1	0,88	0,84	
7	224	174	103	231	180	108	209	161	96	247	231	190	251	233	187	236	211	161	1	0,78	0,46	1	0,92	0,73	1	0,85	0,60	
8	100	74	73	104	79	77	97	73	72	156	105	105	170	120	119	149	102	102	1	0,75	0,74	1	0,69	0,69	1	0,72	0,71	
9	136	93	64	141	96	65	130	89	63	188	148	98	193	156	105	155	114	74	1	0,68	0,50	1	0,73	0,51	1	0,71	0,51	
10	112	87	70	120	91	73	112	84	69	117	81	67	119	83	68	109	75	63	1	0,68	0,47	1	0,78	0,52	1	0,73	0,49	
11	156	110	88	198	144	110	123	89	75	215	172	135	241	208	170	208	160	124	1	0,76	0,62	1	0,69	0,57	1	0,73	0,60	
12	116	71	64	119	72	63	113	69	72	181	120	81	200	136	92	171	113	92	1	0,72	0,57	1	0,81	0,65	1	0,77	0,61	
14	180	127	73	188	133	75	167	119	68	210	155	85	219	167	93	193	141	78	1	0,61	0,57	1	0,67	0,48	1	0,64	0,53	
13										140	104	81	149	111	89	136	102	80		1	0,71	0,40	1	0,74	0,41	1	0,73	0,41
14										191	140	105	212	168	130	188	136	99				1	0,75	0,59				
15										166	136	128	208	182	171	132	109	103				1	0,75	0,57				
																						1	0,84	0,79				

TABLE IV

EXAM. PLE N°	IONIC FORM (Na SALT) - RGB VALUES						NONIONIC FORM - RGB VALUES						R:G:B RATIO			R:G:B RATIO			R:G:B RATIO		
	MEDIUM			LIGHT SPOT			DARK SPOT			MEDIUM			LIGHT SPOT			DARK SPOT			R		
	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B	R	G	B
16	70	59	56	75	60	57	63	54	52	108	86	68	139	110	84	94	77	62	1	0,83	0,79
17	112	84	68	117	88	70	98	73	61	151	115	87	169	128	96	141	107	82	1	0,75	0,61
18	88	65	52	98	72	58	81	61	48	128	99	70	147	112	68	117	89	67	1	0,74	0,59
19	131	102	69	144	110	74	120	94	64	177	143	105	195	162	124	162	130	97	1	0,77	0,52
20	180	145	99	185	149	99	176	140	93	223	189	145	227	199	159	215	183	141	1	0,8	0,54
21	73	63	59	79	67	62	63	56	54	125	100	80	159	128	99	106	85	69	1	0,87	0,81
22	151	112	78	156	116	80	136	102	72	223	177	118	231	191	135	211	166	112	1	0,74	0,52
23	94	68	53	104	75	58	90	65	46	133	100	68	165	127	85	117	88	62	1	0,72	0,55
24	97	64	54	105	70	59	93	63	53	162	115	81	183	130	93	145	102	72	1	0,67	0,56
25	110	69	64	116	72	65	105	65	60	162	104	73	171	112	80	147	93	67	1	0,62	0,57
26	112	80	51	119	86	56	107	77	49	154	114	72	165	124	75	147	109	69	1	0,72	0,46
27	90	64	64	103	73	70	68	54	56	137	94	87	156	105	96	128	90	88	1	0,73	0,73
28	96	66	65	109	76	72	90	64	61	130	88	83	159	113	108	78	66	65	1	0,70	0,67
29	103	73	59	105	73	59	100	70	57	73	55	51	83	63	56	72	55	51	1	0,70	0,57
30										50	46	42	88	80	78	32	28	24			
31										72	66	64	76	70	67	64	58	56			
32										106	104	104	112	109	106	93	90	88			

R:G:B RATIO			R:G:B RATIO			R:G:B RATIO		
IONIC FORM			NONIONIC			AVERAGE		
R	G	B	R	G	B	R	G	B
1	0,83	0,79	1	0,8	0,63	1	0,82	0,71
1	0,75	0,61	1	0,76	0,57	1	0,75	0,59
1	0,74	0,59	1	0,77	0,52	1	0,75	0,56
1	0,77	0,52	1	0,81	0,61	1	0,79	0,57
1	0,8	0,54	1	0,86	0,67	1	0,83	0,6
1	0,87	0,81	1	0,8	0,64	1	0,83	0,72
1	0,74	0,52	1	0,8	0,55	1	0,77	0,53
1	0,72	0,55	1	0,76	0,52	1	0,74	0,53
1	0,67	0,56	1	0,71	0,5	1	0,69	0,53
1	0,62	0,57	1	0,64	0,46	1	0,63	0,51
1	0,72	0,46	1	0,74	0,46	1	0,73	0,46
1	0,73	0,73	1	0,69	0,64	1	0,71	0,69
1	0,70	0,67	1	0,73	0,70	1	0,71	0,68
1	0,70	0,57	1	0,76	0,69	1	0,73	0,63
			1	0,91	0,85			
			1	0,92	0,88			
			1	0,97	0,96			

Biological Example 1 - Inhibition of direct lipoperoxidation on LDL

The inhibition activity of the synthetic vegetal melanins has been carried out according to Visioli F. and Galli C. "Evaluating oxidation processes in relation to cardiovascular disease: a current review of oxidant/antioxidant methodology." in

5 Nutr. Metab. Cardiovasc. Dis. 7: 459-466 (1997).

Human LDL (d=1.021-1.063) were isolated by sequential ultracentrifugation from plasma obtained from healthy, normolipidemic volunteers. Before initiation of the experiments, LDL samples were desalted by size-exclusion chromatography and their protein content was determined according to Lowry, O., et al., J. Biol. Chem.,

10 1951, 193, 265 (1951).

Then 5 mg of the vegetal melanins were added and oxidation started by the addition of the free radical generator 2,2'-azo-bis(2-amidinopropane) dihydrochloride (AAPH) at concentrations of 5 mM. Incubation were carried out at 37 °C in a shaking bath and aliquots were withdrawn at different times for the analysis of the

15 tiobarbituric acid reacting substance (TBARS) as oxidation marker, as shown in Table II.

TABLE II

Inhibition of LDL oxidation from 5 mM AAPH by vegetal melanins

	Vegetal melanin	nMol TBARS/ mg LDL	Inhibition (%)
20	Control (no melanin)	32.54	0
	of the Example 16	21.35	34%
	of the Example 17	25.23	22%
	of the Example 1	30.22	7%
	of the Example 19	30.14	7%
25	of the Example 8	21.87	33%
	of the Example 31	22.01	23%

These data shown that the radical quenching activity is higher for the galloylated vegetal melanin of the Example 8 s well as in the "mixed type" vegetal melanin, thus which contain eumelanin precursors and open-ring polyphenols, either obtained by

30 chemical or enzymatic synthesis, as for Example 16, 17 and 31. The lower value are reached by the presence of the unstable flavonol moiety from quercetin, as in

Examples 1 and 19.

Biological Example 2 - Dose-dependent inhibition of direct lipoperoxidation of LDL

The test was carried out as described in the Biological Example 2 but using the vegetal melanin of Example 16 at different concentrations. Results are listed in Table

5 III.

TABLE III

Concentration (ppm)	nMol TBARS/ mg LDL	Inhibition (%)
0	7.05	-
1 ppm	4.83	31%
10 5 ppm	3.57	49%
10 ppm	2.42	66%
20 ppm	1.92	73%

As it can be noted, the radical scavenging activity (primary antioxidant inhibition) of the vegetal melanin follows an approximately first order reaction, with a significantly
15 high decrease of the lipoperoxidation behavior for small increase of the vegetal melanin.

Biological Example 3 - Inhibition of metal-dependent peroxidation of LDL

The test was carried out according Visioli F. and Galli C. (ibid): LDL were separated by sequential ultracentrifugation from human plasma and the samples were prepared
20 by diluting LDL in PBS at a final concentration of 200 ug protein/ml, to a final concentration of 200 ug protein/ml. The probes were thereby pre-incubated for 30 minutes at 37°C with 5 mg of the vegetal melanins under investigation, then copper sulfate (5 uM, final concentration) was added to start the oxidation, which was carried out at 37°C in a shaking bath. At the end of the incubation period, samples
25 were withdrawn for the quantitation of the tiobarbituric acid reacting substances (TBARS). Results are listed in Table IV.

TABLE IV

Vegetal melanin	nMol TBARS/ mg LDL	Inhibition (%)
Control (no melanin)	58.97	0
30 of the Example 16	11.22	81%
of the Example 17	22.33	62%

of the Example 1	45.59	23%
of the Example 19	47.54	19%
of the Example 8	12.93	78%
of the Example 31	45.10	24%

- 5 The aforelisted data exhibit a high correlation between the potential chelating activity and the structure of vegetal melanin. Thus, the chelating behaviour is significantly high in all vegetal melanins but those formed with quercetin among starting monomers, still retaining an appreciable secondary antioxidant activity.

Biological Example 4 - Dose-dependent inhibition of metal-dependent

10 lipoperoxidation

The test was carried out as described in the Biological Example 3 but using the vegetal melanin of Example 16 at different concentrations. Results are listed in Table III.

TABLE V

15	Concentration (ppm)	nMol TBARS/ mg LDL	Inhibition (%)
	0	10.15	-
	1 ppm	8.87	13%
	5 ppm	4.91	52%
	10 ppm	1.39	86%
20	20 ppm	1.43	86%

- The figures show that the inhibition by vegetal melanin of the metal-induced peroxidation does follow a steep slope, with almost complete deactivation already at 10 ppm. Noteworthy, this antioxidant activity is higher than most plant polyphenols, i.e. the vegetal melanin monomers. Not surprisingly, the concentration of 20 ppm do
- 25 not offer any further protection, as the copper ions are already fully chelated by a polydentate complex with the vegetal melanin, thus any Fenton-like activity is therefrom inhibited.

Applicative Examples 1-3 - Foundation

100 g of each foundations contain:

30		Appl. Ex. 1	Appl. Ex. 2	Appl. Ex. 3
	Triethanolamine stearate	2.5 g	2.5 g	2.5 g

	Glycerol mono- e distearate	0.4 g	0.4 g	0.4 g
	Magnesium silicate	1.9 g	1.9 g	1.9 g
5	Pigment formed with the vegetal melanin of Example 13 (*)	14.0 g	10.0 g	3.7 g
	Pigment formed with the vegetal melanin of Example 16 (*)	2.9 g	8.0 g	-
10	Pigment formed with the vegetal melanin of Example 28 (*)	-	2.1 g	15.0 g
	Titanium dioxide (rutile)	10.0 g	10.0 g	10.0 g
	Miscela di PEG-6 e PEG-32	9.0 g	9.0 g	9.0 g
15	Nylon micronized	9.0 g	9.0 g	9.0 g
	Cyclomethicone	13.0 g	13.0 g	13.0 g
	Propylene glycol	5.0 g	5.0 g	5.0 g
20	Glycerine	4.5 g	4.5 g	4.5 g
	Preservatives	0.5 g	0.5 g	0.5 g
	Deionized water	qb to 100 g	qb to 100 g	qb to 100 g
25	(*) In 100 ml of the solution of the vegetal melanin are suspended 13 g of alumina, the slurry is stirred and added with a solution of AlCl ₃ 1M until pH 6-7. The slurry is then washed with demineralized water, filtrated, then dried in nitrogen stream. and the pigment is finally grinded.			
30	The fatty phase containing the oils and stearic acid and the aqueous phase containing triethanolamine are separately heated to 80 °C. The mixture is emulsified at 80°C and cooled slowly. During the cooling, the mixture of pigments, previously ground in			

propylene glycol and cyclomethicone are added.

Applicative Examples 4-6 - Preparation of a suntanning lotion

100 g of each suntanning lotions contain:

		Appl. Ex. 4	Appl. Ex. 5	Appl. Ex. 6
5	EO-21-stearyl ether (ICI G-1800)	5.0 g	5.0 g	5.0 g
	Isopropyl myristate	10.0 g	10.0 g	10.0 g
	Vegetal melanin solution of Example 4	1.0 g	1.0 g	1.0 g
	Vegetal melanin solution of Example 6	1.0 g	1.0 g	1.0 g
	Vegetal melanin solution of Example 14	1.0 g	1.0 g	1.0 g
10	Oleyl alcohol	2.0 g	2.0 g	2.0 g
	2-Hydroxy-3,3,5-trimethylhexyl ester of benzoic acid	4.0 g	4.0 g	4.0 g
	BHA	0.05 g	0.05 g	0.05 g
	Perfume	0.3 g	0.3 g	0.3 g
15	Distilled water	q.b. to 100 g	q.b. to 100 g	q.b. to 100 g

The mixture is heated to 70°C and preheated distilled water, and the vegetal melanin is added thereto. The resultant mixture is stirred and allowed to cool to room temperature.

Applicative Example 6 and Comparative Example 1 - Protective-tanning creams with

test of suppression of UV-induced erythema

100 g of each emulsions contains:

		Appl. Ex. 6	Comp. Ex. 1
	Cetostearyl alcohol	3.0 g	3.0 g
	Oleic polyoxyethylen glyceride	5.0 g	5.0 g
25	Gliceryl monostearate	4.0 g	4.0 g
	2-Ethylexyl cocoate	2.0 g	2.0 g
	Tocopheryl acetate	0.5 g	0.5 g
	Ascorbyl palmitate	0.1 g	0.1 g
	Phospholipids (soy lecithin 63%)	3.0 g	3.0 g
30	Vegetal melanin solution of Example 25	3.0 g	-
	Caramel solution 10% w/v in NaOH 1N	-	3.0 g

Preservatives	0.3 g	0.3 g
Deionized water	qb to 100 g	qb to 100 g

The mixture was heated to 70°C and a preheated liposomal phymelanins, obtained by dispersing at 20.000 rpm for 10 minutes the solutions of
 5 vegetal melanin with soybean lecithin and water, is added thereto. The same procedure but with a 10% solution of caramel in NaOH 1N is carried out in Comparative Example 1.

The resultant mixtures are stirred and allowed to cool to room temperature.

Test of suppression of UV-induced erythema by vegetal melanin

10 A comparative study on vegetal melanin radiation protectant activity on the suppression of UV-induced erythema is assessed according the method described by Bangha E; Elsner P; Kistler GS (Arch. Dermatol. Res., 288(9):522-6. 1996).

3 volunteers were topically treated by applying approximately 10 g of the composition of Example 6 in the right side of the back and 10 g of the composition
 15 without vegetal melanin (as per Comparative Example 1) on the controlateral side, then irradiated with 0.1 J/cm² UVB on two 5-cm² areas on the lower back.

The UV-induced erythema was examined 24 h after irradiation by visual scoring. The differences in redness is shown in Table III.

TABLE III

20 Visual scoring by applying topical compositions with and without vegetal melanin
 Redness after 24 hours

Comparative Example 1 (right side, no vegetal melanin) persistant

Example 14 (left side, with vegetal melanin) none

As it can be noted, the presence of vegetal melanin contributes to the prevention of
 25 UV-induced erythema.

Applicative Example 7 – Oral composition with vegetal melanin

A composition is prepared in the form of ampoules with the following ingredients:

Vegetal melanin solution of Example 11	2.5 g
30 ascorbic acid	12.0 mg
maltol	30.0 mg

L-tartaric acid 15.0 mg

strawberry flavor 2.0 mg

sodium citrate 1.0 g

solubilizing the mixture in distilled water qb to 250 ml.

- 5 It should be understood that the specific forms of the invention herein illustrated and described are intended to be representative only. Changes, including but not limited to those suggested in this specification, may be made in the illustrated embodiments without departing from the clear teachings of the disclosure.
- Accordingly, reference should be made to the following appended claims in
- 10 determining the full scope of the invention.

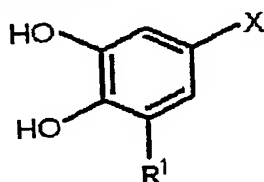
CLAIMS

1. Synthetic vegetal melanins obtainable by in vitro polymerization of at least one monomer unit (a), and optionally at least one compound (b), wherein:
- (a) is a plant polyphenol; and

5 (b) is an eumelanin precursors;

said synthetic vegetal melanin being characterized by a Red:Green:Blue ratio comprised between 1 : 0.88 : 0.84 and 1 : 0.64 : 0.41.

2. Synthetic vegetal melanins according to claim 1, wherein the plant polyphenol is a vegetal substance of formula (I):



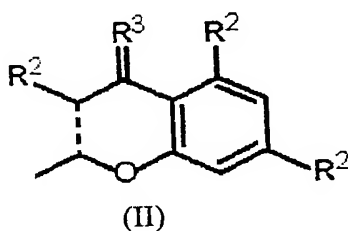
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(I)

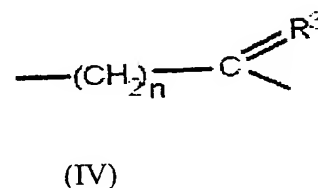
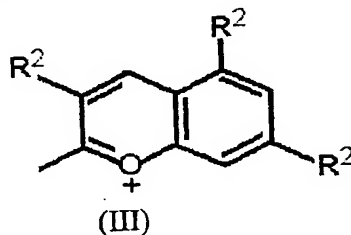
wherein:

R^1 represents hydrogen, hydroxy, or methoxy group; and

X represents a residue of formulas (II), (III) or (IV)



15



wherein:

n is 0, 1 or 2;

R^2 represent, each independently, hydrogen, hydroxy, methoxy, or an O-glycosylated group;

20 R^3 represents, one oxygen, two hydrogen atoms, or one hydrogen and one hydroxy group; and

the phantom line between C_2 - C_3 represents a double or a single bond, as well as salts, esters and solvates thereof.

- 25 3. Synthetic vegetal melanins according to claims 1 to 2, wherein the plant

polyphenol of formula (I) with X being a residue of formula (II) is a flavonoid selected in the group consisting in quercetin, fisetin, fustin, luteolin, OPC, catechin, epicatechin, GC, GCG, EGC, EGCG, myricetin, dihydroquercetin and mixture thereof.

- 5 4. Synthetic vegetal melanins according to claims 1 to 2, wherein the plant polyphenol of formula (I) with X being a residue of formula (III) is a flavonoid anthocyanins selected in the group consisting in cyanidin, delphinidin and mixture thereof.
- 10 5. Synthetic vegetal melanins according to claims 1 to 2, wherein the plant polyphenol of formula (I) with X being a residue of formula (IV) is an open ring dihydroxyphenol selected in the group consisting in hydroxytyrosol, protocatechuic acid, protocatechuic aldehyde, gallic acid, tannic acid and mixture thereof.
- 15 6. Synthetic vegetal melanins according to claims 1 to 2, wherein the eumelanin precursor is selected in the group consisting in L-dopa, DHI, DHICA, dopamine, pyrocatechol, pyrogallol and mixture thereof.
- 20 7. Method of producing synthetic vegetal melanins according to claims 1 to 6 comprising bubbling air or oxygen through an alkaline aqueous solution of the monomers at pH 10 or higher, during 12 to 48 hours, at a temperature ranging from 10 to 90°C.
8. Method of claim 7, further comprising catalytic amounts of pro-oxidant metals selected in the group consisting in Cu^{++} , Fe^{++} , Ni^{++} , Co^{++} and mixture thereof.
- 25 9. Method of claim 7 where the chemical oxidizing agent is selected in the group consisting in hydrogen peroxide, hydrogen iodide, ammonium persulfate, potassium permanganate, magnesium perchlorate and mixture thereof.
- 30 10. Method of producing synthetic vegetal melanins according to claim 1 to 6 comprising bubbling air or oxygen through an buffered aqueous solution of the monomers at pH from 9.5 to 4.5, during 12 to 48 hours, at a temperature ranging from 20 to 45°C in presence of a suitable enzyme.
11. Method of claim 10, wherein the enzyme is selected in the group consisting in tyrosinases, polyphenoloxidases (catagchol oxidases), phenolases.

(phenoloxidas), peroxidases, laccases, lipoxygenase, and mixture thereof.

12. Method according to claims 10 to 12 wherein the monomer units (a) or (b) are formed in situ from the pre-monomers, bearing a monophenolic moiety.
13. Method of claim 12 where the pre-monomers are selected in the group consisting in dihydrokaempferol, armadendrin, p-hydroxybenzaldehyde, PHBA, tyrosol, p-coumaric acid, apigenin, kaempferol, pelargonin, genistein, tyrosine, tyramine, 5-hydroxy-indole and mixture thereof.
14. Cosmetic composition comprising as active ingredient at least one synthetic vegetal melanins according to claims 1 to 6.
15. Cosmetic composition according to claim 14, for facial make-up, hair dyes, tanning, anti-sun, toiletry, moisturizing and protecting skin.
16. Pharmaceutical or nutritional composition having anti-inflammatory and immunomodulation activity which comprises as active ingredient at least a synthetic vegetal melanin according to claims 1 to 6.

FIGURE 1

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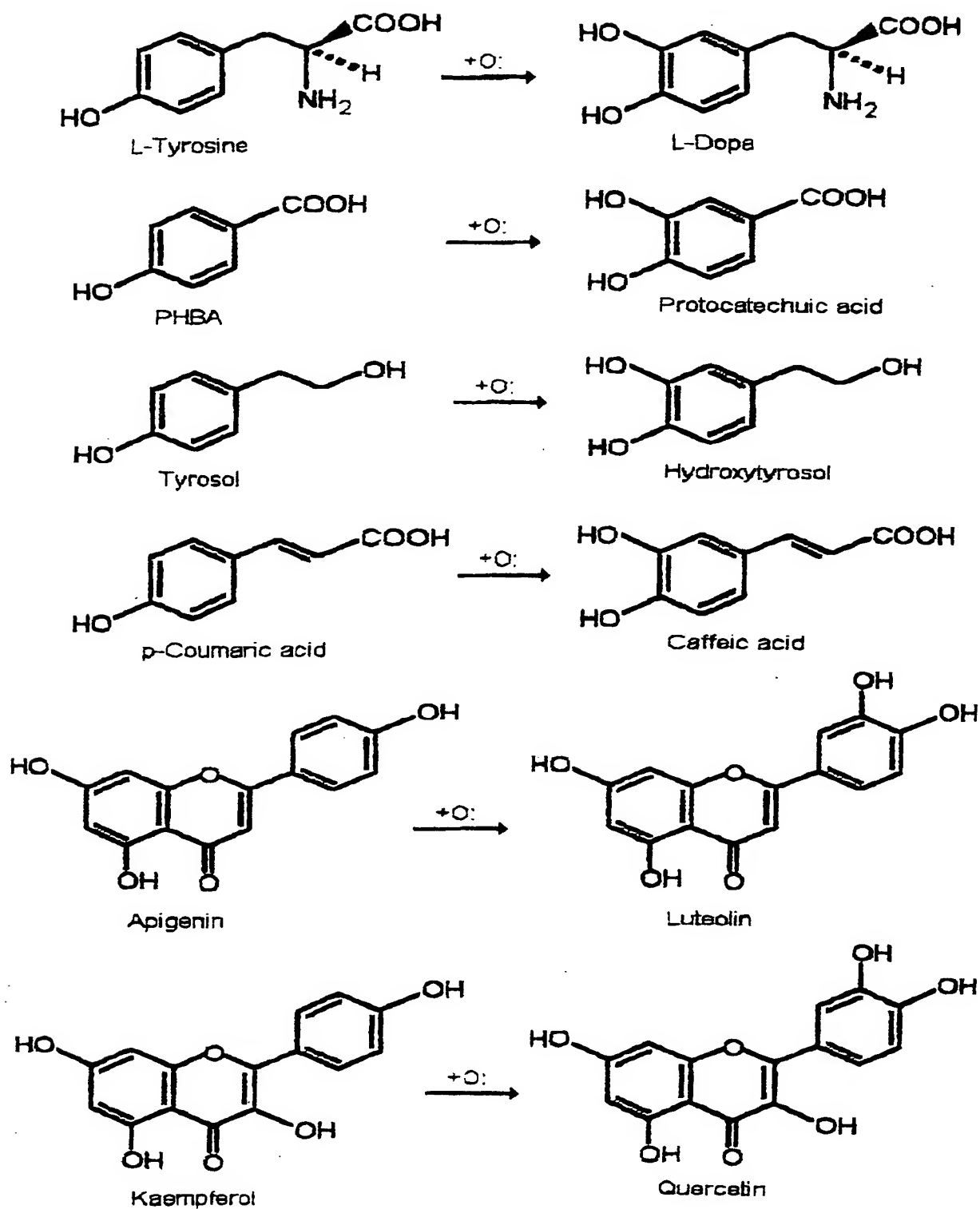


FIGURE 2

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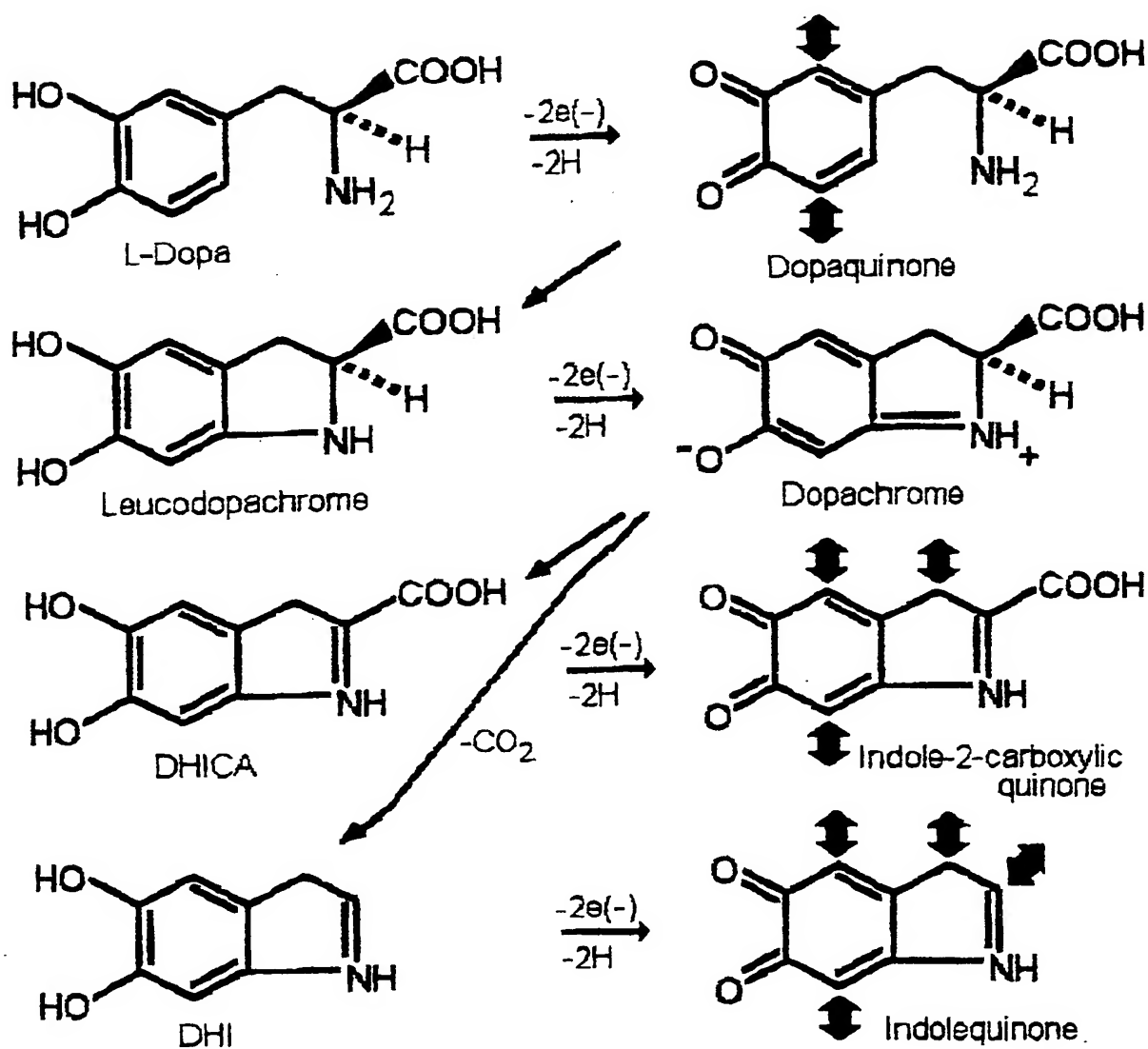
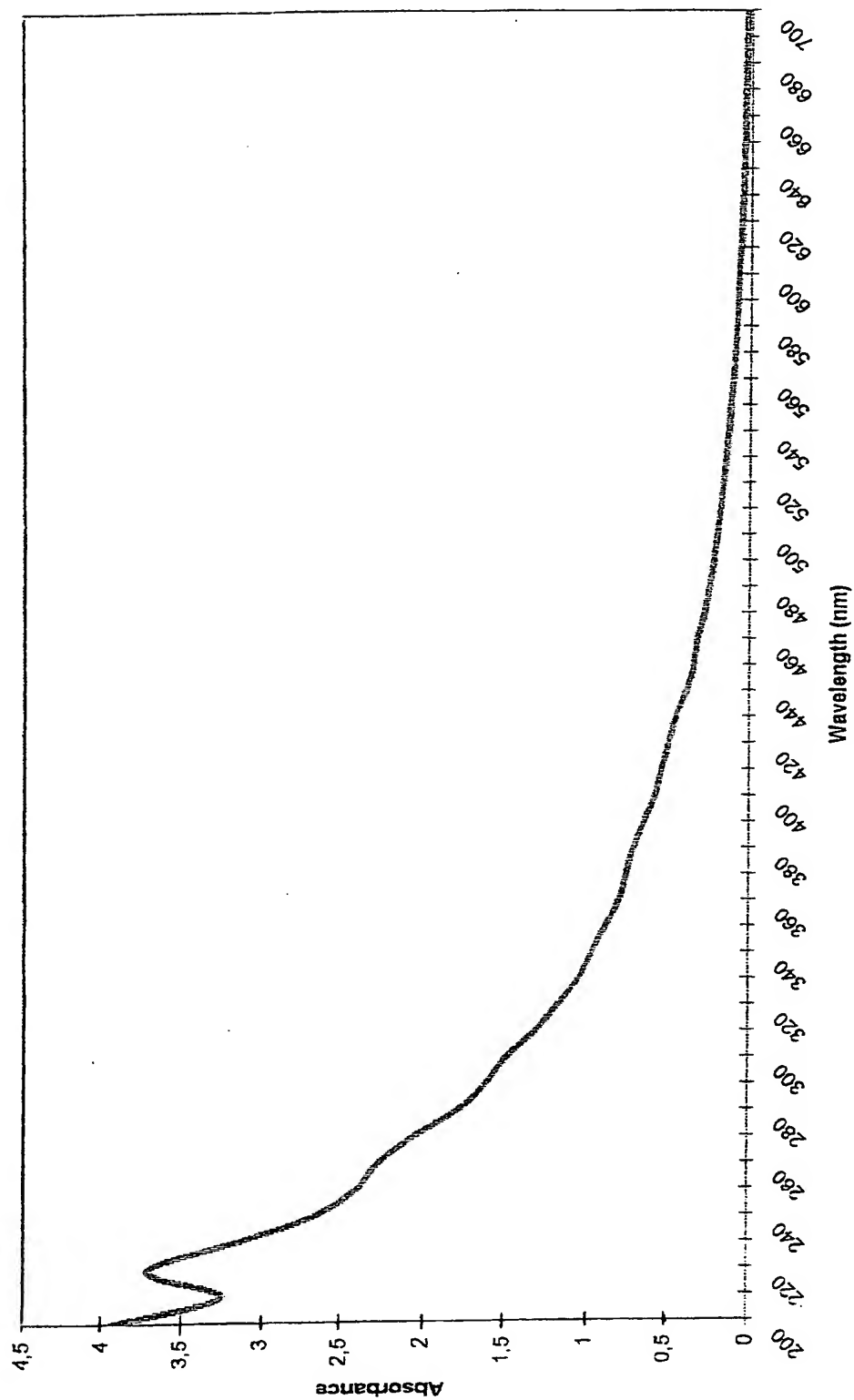


FIGURE 3
Example 15



Example 17

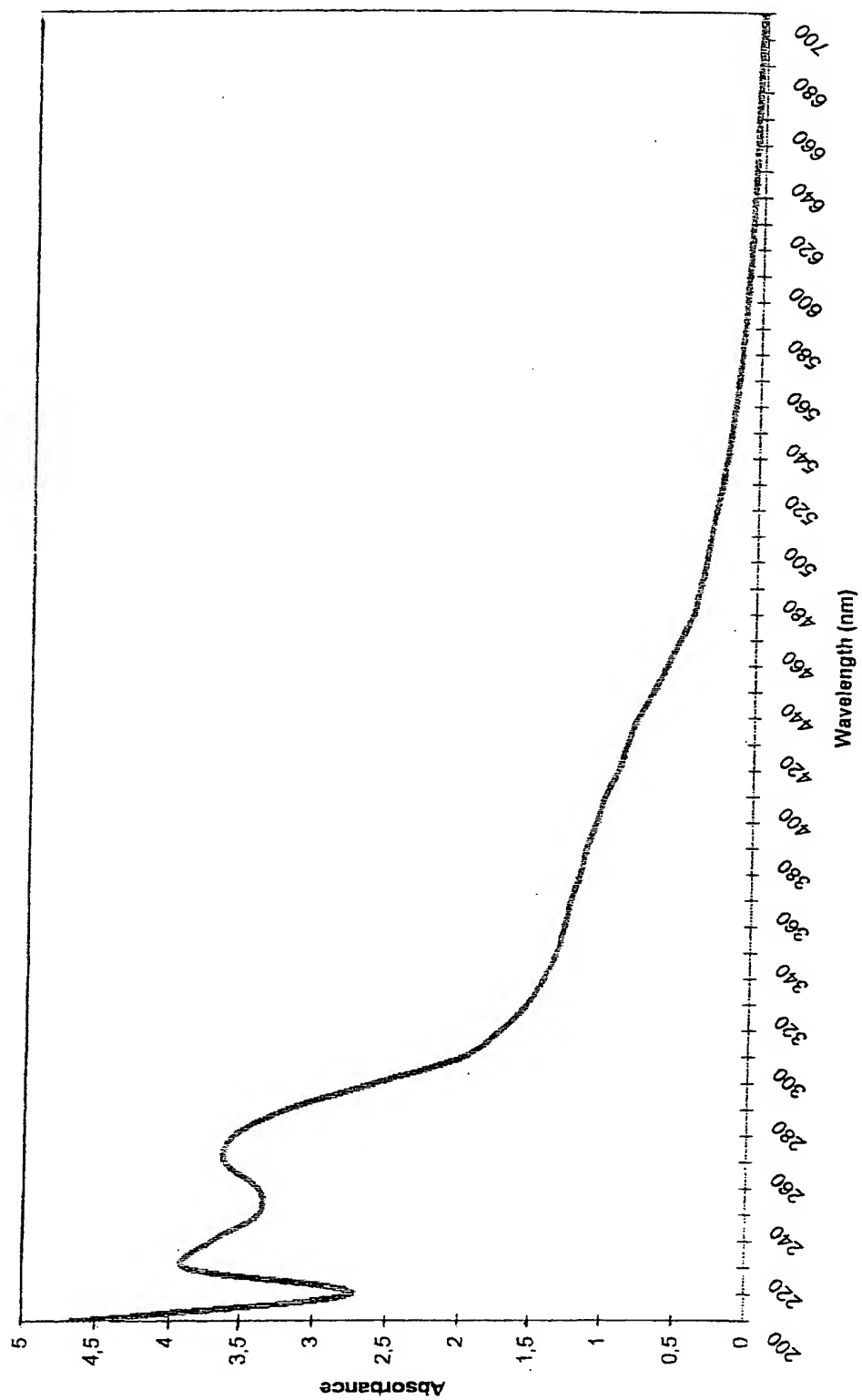


Figure 4

Example 18

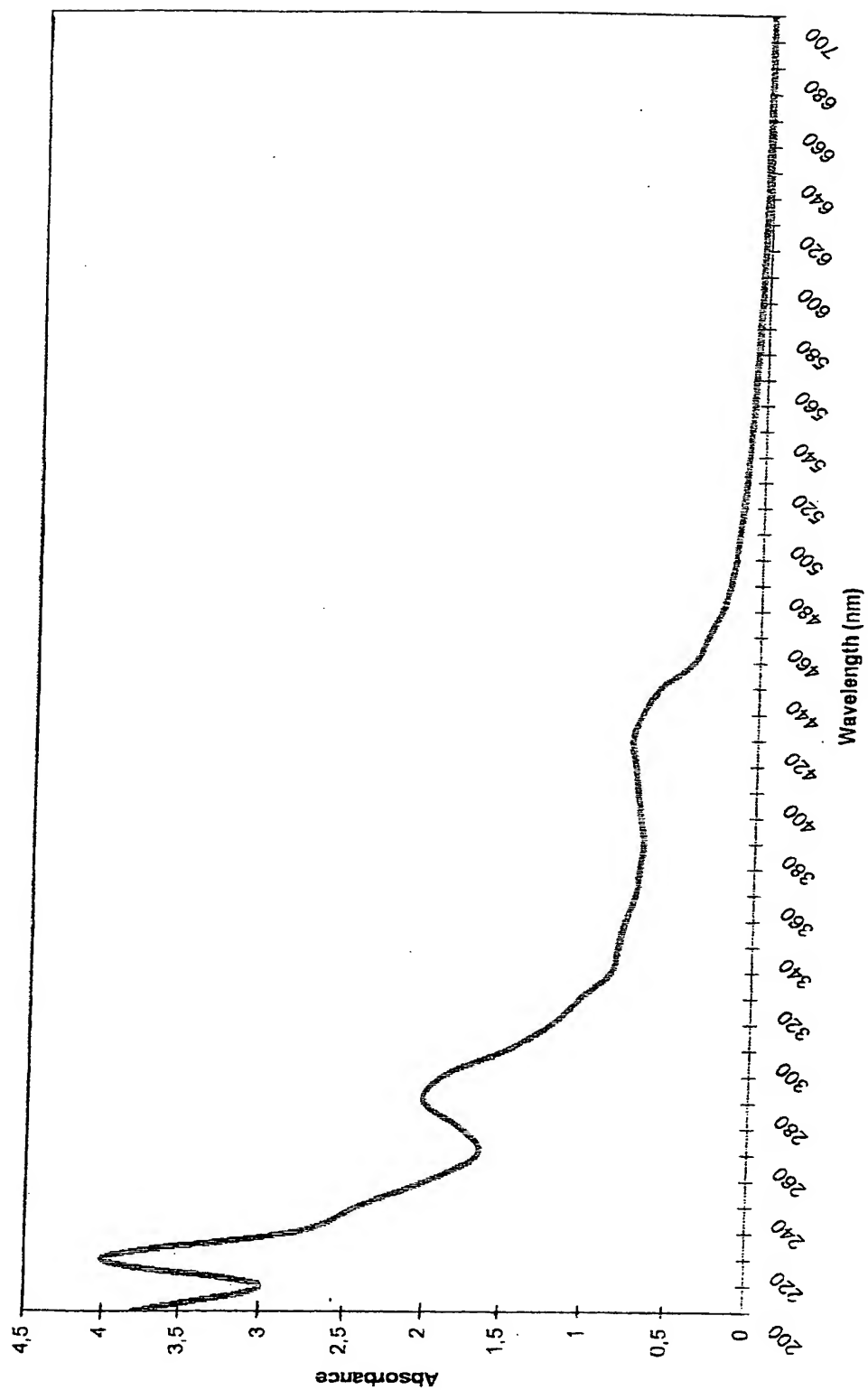
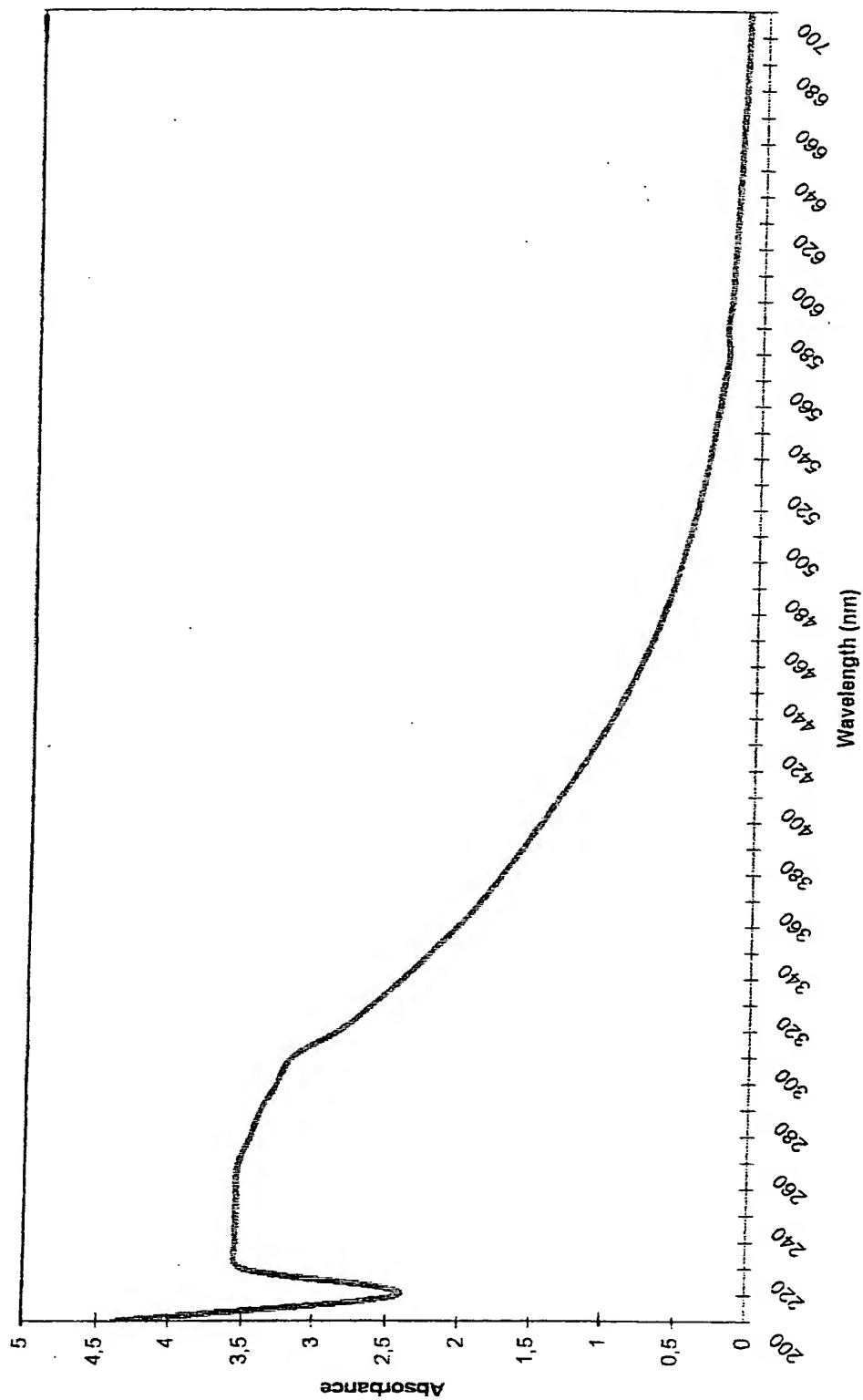


Figure 5

Example 23

Figure 6



INTERNATIONAL SEARCH REPORT

International Application No
PCT/IB 00/01276

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C09B61/00 C09B69/10 A61K7/42

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C09B A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

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	-/-	

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- *&* document member of the same patent family

Date of the actual completion of the international search

21 December 2000

Date of mailing of the international search report

02/01/2001

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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